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Unravelling the life history of Amazonian fishes through otolith microchemistry

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Amazonian fishes employ diverse migratory strategies, but the details of these behaviours remain poorly studied despite numerous environmental threats and heavy commercial exploitation of many species. Otolith microchemistry offers a practical, cost-effective means of studying fish life history in such a system. This study employed a multi-method, multi-elemental approach to elucidate the migrations of five Amazonian fishes: two 'sedentary' species (Arapaima sp. and Plagioscion squamosissimus), one 'floodplain migrant' (Prochilodus nigricans) and two long-distance migratory catfishes (Brachyplatystoma rousseauxii and B. filamentosum). The Sr: Ca and Zn: Ca patterns in Arapaima were consistent with its previously observed sedentary life history, whereas Sr: Ca and Mn: Ca indicated that Plagioscion may migrate among multiple, chemically distinct environments during different life-history stages. Mn: Ca was found to be potentially useful as a marker for identifying Prochilodus's transition from its nursery habitats into black water. Sr: Ca and Ba: Ca suggested that B. rousseauxii resided in the Amazon estuary for the first 1.5-2 years of life, shown by the simultaneous increase/decrease of otolith Sr:Ca/Ba:Ca, respectively. Our results further suggested that B. filamentosum did not enter the estuary during its life history. These results introduce what should be a productive line of research desperately needed to better understand the migrations of these unique and imperilled fishes.

1. Introduction

Otoliths, the calcified ear-stone organs in fish hearing/balance systems, have been widely used to understand fish life history. Otolith incremental deposition forms the basis of age and growth

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studies [1–3], and incorporated elements and isotopes provide additional information on the life history of an individual [4–6]. Otoliths are metabolically inert, recording information over a fish's entire life [5]. Most studies on the chemical composition of otoliths have focused on coastal marine and diadromous fishes due to the great chemical disparity of the environments exploited by these fishes (e.g. river and estuarine versus marine environments). Otolith microchemistry studies (using micro-and trace elements) in freshwater fishes have elucidated natal origins [7–9], migration patterns [10–12] and pollution exposure history [13,14], providing critical information needed for the management of freshwater fisheries [15]. However, little work has been done on otolith microchemistry in tropical freshwater systems, even though they are under increasing anthropogenic pressures.

There is great need to study the life history of commercial fishes in the Amazon and other large South American basins, as there are dozens of exploited taxa with widely varying life histories that are poorly understood [16]. These basins provide ideal systems to study tropical fish otolith microchemistry. The Amazon Basin is geochemically diverse and complex, so many otolith chemical markers may be present. Despite the potential for otolith microchemistry studies, only five studies have used microchemical markers to understand fish migrations in the Amazon [17–21]. These studies used multicollector-inductively coupled plasma mass spectrometry (ICPMS), a powerful but expensive tool for quantifying isotope ratios, limited in the number of elements that can be sampled in one ablation pass. While Sr isotope ratios have proved to be useful in freshwater systems, it is well established that otoliths entrain many other useful micro- and trace chemicals in freshwaters, including manganese [22], bromine [23] and selenium [13], among others. An approach employing multiple elements to the study of fish otolith microchemistry could help elucidate the life histories and ecology of Amazonian fishes.

Low-resolution laser ablation–ICPMS (LA–ICPMS) and scanning X-ray fluorescence microscopy (SXFM), though unsuited for accurate quantification of multiple Sr isotopes, offer another cost-effective way to sample a multitude of elements simultaneously. The two methods complement one another in their ability to measure particular elements that the other cannot (e.g. LA–ICPMS can accurately measure Ba, while SXFM can measure Se) and may complement data provided by multicollector–ICPMS [24]. The flexibility to sample so many elements simultaneously is particularly important in poorly studied systems, like the Amazon, where elements that may be useful markers are largely unknown. Otolith microchemistry also provides a stark contrast to other methods of studying fish migration. Mark–recapture studies are prohibitively expensive in large or remote systems [25], while fish in tagging studies are often intensely harvested by fishermen (e.g. [26]) or migrate distances too great to study with these methods [27].

Here, we demonstrate the usefulness of a multi-element approach based on low-resolution LA– ICPMS and SXFM for unravelling the life histories of Amazonian commercial fishes.

1.1. Life histories of Amazonian fishes

The Amazon Basin is a complex network of interconnected terrestrial and aquatic environments, with chemical diversity dictated by the bedrocks and soils through which river tributaries pass [28]. The most abundant fish populations are in the várzea floodplains, which experience 10 m seasonal inundations by the sediment-laden and nutrient-rich whitewater rivers originating in the Andes Mountains [29,30]. When water levels rise, the main river and floodplains become hydrologically interconnected, prompting fishes to migrate out of lakes and river channels and onto floodplain forests and floating meadows (i.e. grasses) [31]. The flooded forests and floating meadows provide diverse and abundant food sources such as periphyton, insects and fruits, and they act as vital nursery habitats for the larvae and juveniles of many species [32–34]. As flood waters recede, most fishes migrate back to main river channels, where temperature and oxygen levels are more stable and tolerable, although many species adapted to warm and/or hypoxic conditions migrate to floodplain lakes and smaller channels during low waters where water quality is generally poor [35].

In the estuary of the Amazon, about 300 commercially important fish species [36] are supported by biological productivity that is driven not only by rising and receding floodwaters but also by tidal movements [37,38]. Some large migratory catfish species are hypothesized to use the estuary as nursery grounds [37], although a recent study suggests that the life histories of these fishes may be variable within and among related species [17].

Three main migratory strategies are recognized among Amazonian commercial fishes to maximize the spatial and temporal heterogeneity of resource availability. 'Sedentary' fishes generally live in floodplain lakes and migrate laterally through floodplain habitats following flood pulses, a behaviour documented in detail in the giant *Arapaima* sp. [39]. Sedentary fishes are typified by *Plagioscion squamosissimus*, a

species common to floodplain lakes [40,41]. Studies of the larval ecology of *Plagioscion* indicate that they tolerate diverse water conditions, but prefer spawning in lentic areas with high temperature, low pH and low dissolved oxygen [42,43]. There is little work on the migration of adult individuals (but see [44,45]). It can be expected that *P. squamosissimus* migrates through floodplain habitats throughout the hydrological cycle like other sedentary fishes.

'Floodplain migratory' fishes are typified by various characoid fishes, such as Prochilodus nigricans. Available studies indicate that adult Prochilodus migrate upwards of several hundred kilometres into, or upstream in, the whitewater rivers of the Amazon mainstem (or major tributaries) as waters begin to rise [33,46]. Large schools broadcast spawn in these whitewater rivers, after which adults migrate laterally onto floodplain habitats to feed. Eggs and larvae drift downriver and settle in the flooded forest or floodplain meadows, where the developing juveniles feed, grow and, eventually, migrate back upstream at an age of about six months to complete the cycle [47]. By migrating between the nutrientrich white water and nutrient-poor, acidic black waters, Prochilodus are exposed to more varied water chemistry than most other Amazonian fishes. However, the migration patterns of Prochilodus species and populations apparently vary in direction and timing in different Amazon tributaries or South American rivers (for summary, see [33]). The migratory ecology of *P. nigricans* in the Ecuadorian Amazon remains poorly studied. Silva & Stewart [48] found that growth parameters for P. nigricans in the Ecuadorian Amazon were inconsistent with those in Bolivia or Central Amazon (i.e. forming annuli versus biannuli in the latter two regions). They suggested a shorter hydrological cycle as a possible factor in the lower growth rates that they observed. This suggests that the time at which juveniles first migrate to rejoin adult populations might also differ among populations.

'Long-distance catfish' migrants are typified by several of the large pimelodid catfishes like *Brachyplatystoma rousseauxii* and *B. filamentosum*. Early studies indicated that adults migrate into the headwater rivers along the Andean foothills during rising waters, where they broadcast spawn [27,49,50]. Eggs, larvae and small juveniles drift downriver for thousands of kilometres, with some of them reaching the estuary. Here the small juveniles settle, feed and grow for several years before migrating up the Amazon mainstem towards the headwaters. The distance migrated during the complete cycle may total upwards of 8000 km [21]. However, the extent to which these fishes use the estuary as a nursery ground is unclear. Recent studies analysed strontium isotope ratios in otoliths and showed that these fishes have varied life histories, both within and among species [17,21]. Some individuals appear to home back to natal spawning grounds [21], indicating more complex life histories than previously thought.

Active research and funding for studying the life histories of Amazonian fishes are minimal, despite the commercial value of these fishes and the multiple environmental threats they face. Overfishing has led to considerable declines in some species [51], while deforestation is producing stronger, earlier flood pulses, altering the flood cycle that migratory fishes rely upon [52]. Hydroelectric damming, long considered a threat to Amazonian migratory fishes [37], is widespread, blocking migration routes and disrupting flood pulses [53,54]. Installation of 'fish ladders' to aid migratory fishes in circumventing these obstructions has been ineffective [55–58]. Pollution from gold mining [59], agriculture [60] and oil exploration [61] is damaging water quality, affecting the health of fish and humans [62–64]. Many of these species, particularly the migratory catfishes, traverse international boundaries during spawning migrations, which further complicates efforts to study their biology or implement effective management strategies. Otolith microchemistry not only reveals the entire recorded life history of these fishes, it also affords a means of doing it cost-effectively.

Here, we asked the following research questions regarding LA–ICPMS and SXFM. (i) Can these methods be used to identify (small-scale) migrations in sedentary fishes? (ii) Can they identify shifts between life-history stages in floodplain migrants? (iii) Can they identify habitat use in juvenile long-distance migrants? This study identified new chemical markers in the otoliths of five Amazonian fishes and unravelled new information on their life histories.

2. Material and methods

This study investigated the migratory behaviours of two sedentary species (*Arapaima* sp. and *P. squamosissimus*), one floodplain migrant (*P. nigricans*), and two long-distance migratory catfishes (*B. rousseauxii* and *B. filamentosum*) by analysing chemical patterns in their otoliths using low-resolution LA–ICPMS and SXFM. LA–ICPMS provided data collected along transects on the otoliths. This method can quantify barium, an important trace element in otolith studies [5], which cannot reliably be measured

Table 1. Summary of the sources used to age each species, the ageing method used within the study, whether the study reported one growth mark (annulus) or two (biannulus) per year, and whether the study validated the growth rings at age.

species	ageing method	growth mark	literature source	ring validation
Arapaima sp.	scales	annulus	Watson 2011 ^{a, b} [66]	yes
Plagioscion squamosissimus	otoliths	annulus	Loubens 2003 [40]	yes
Prochilodus nigricans	otoliths	annulus	Silva & Stewart 2006 ^a [48]	no
Prochilodus nigricans	length-frequency	n.a.	Silva 2000 ^{a,b} [47]	n.a.
Brachyplatystoma rousseauxii	otoliths	biannulus	Alonso & Fabré 2003 ^b [67]	yes
B. filamentosum	otoliths	biannulus	Alonso & Fabré 2003 ^{b, c} [67]	yes

^aOtoliths collected for this study were used in this study.

^bThesis or dissertation.

^cAlonso & Fabre [67] studied only *B. rousseauxii*. We applied their methods to *B. filamentosum*.

at low concentrations with SXFM due to measurement interference from a secondary calcium peak. SXFM provided data for elements not sampled by LA–ICPMS (such as selenium), as well as twodimensional chemical 'maps' of entire otolith sections that reveal possible chemical heterogeneity within the otolith structure. We also analysed some otoliths using both methods to serve as a check for elements that can be quantified by both methods. Changes in measured elements over a fish's life were used to infer migration to and/or residency in various habitats.

2.1. Otolith collection and preparation

Fish otoliths were obtained from various locations. One otolith from *Arapaima* sp. (n = 1; 160 cm standard length (S.L.)) was collected in November, 2008 from Inkapati Head Pond near Apoteri, adjacent to the Essequibo River mainstream in Guyana (4°7.027′ N, 58°29.531′ W). Sagittal otoliths from *Plagioscion* (n = 1; length unknown) and lapillae from *Prochilodus* (n = 6; 97–267 mm S.L.) were collected in the Ecuadorian Amazon in April and May, 1999, from the lower reaches of the Cuyabeno drainage (0°15.50′ S, 75°53.97′ W), a blackwater river, near its confluence with the whitewater Rio Aguarico. A lapillus from *B. rousseauxii* (n = 1; 85 cm fork length (F.L.)) was also collected in April, 1999, near the confluence of the blackwater Rio Sabalo and Rio Aguarico (0°23.58′ S, 75°40.083′ W), Ecuador. Additional lapillae from *B. rousseauxii* (n = 1; 118 cm S.L.) and *B. filamentosum* (n = 1; 130 cm S.L.) were collected in August and December 1995, respectively, from the whitewater Rio Caquetá, near Araracuara in Colombia (0°37.0′ S; 72°23.112′ W). All otolith preparations were completed at the State University of New York, College of Environmental Science and Forestry (SUNY–ESF) in Syracuse, NY, USA.

Otoliths were first cleaned of organic matter by immersion in a dilute (10% v/v) bleach-water solution followed by gentle abrasion and rinsing in de-ionized water. Further preparation of otoliths for microchemical analyses was adapted from Secor *et al.* [65]. Large lapillar (*B. rousseauxii* and *B. filamentosum*) and sagittal (*P. squamosissimus*) otoliths were cast into circular moulds using EpoFix (Struers) cold-set epoxy. Otoliths were placed in a dehydrator overnight to allow the epoxy to set. Epoxy blocks were sectioned through the core in the transverse plane with a low-speed diamond saw (Buehler, IsoMet) and then polished using progressively finer grades of aluminium oxide lapping paper until the otolith core was exposed (determined by bright-field light microscopy). This process usually involved several repetitions before the core was fully visible. *P. nigricans* otoliths were small enough that they required no cutting and were instead mounted on fused-quartz glass slides using cyanoacrylate adhesive (Loctite) and polished to the core using the methodology described above. Samples were mounted on clean glass or with no backing at all for SXFM analysis, and were mounted on petrographic slides for LA–ICPMS analysis.

Fish ages were also estimated based on previous studies using otoliths, scales and/or lengthfrequency distributions (table 1). We note, however, that many studies interpreting the ages and/or validating the ages of these fishes remain unpublished or are unavailable. Therefore, we cautiously estimated age for the species herein to provide a broader ecological context for our findings. One notable uncertainty regards the formation of one growth check (annulus) or two growth checks (biannul-) per year. For example, Watson [66] found, via recapture of three individuals, that *Arapaima* sp. scales in Guyana consistently formed one growth mark per year, while Arantes *et al.* [68] verified two marks per year. Silva & Stewart [48] found that *P. nigricans* in Ecuador had annuli, while it appears that the same species may have biannuli in the Central Amazon [69]. For the purposes of this study, we relied on ageing methods developed in the regions from which our specimens came whenever possible. Extensive studies validating otolith ring formation may further elucidate geographical patterns of annuli versus biannuli within and among populations and species. Until such data become available, we must approach our age estimates with a cautious eye.

2.2. Sampling with scanning X-ray fluorescence microscopy

The SXFM was conducted at the F3 Beamline Station at the Cornell High Energy Synchrotron Source (CHESS). Samples were taped to cardboard slide frames, and then clipped in place to secure them for sampling. A double-bounce multilayer monochromator was used to create a 16.1 keV beam with 0.6% bandpass. A single-bounce glass capillary [70,71] focused the X-ray beam to a 25–75 µm (depending on sample size) spot on the sample with a photon flux of approximately 1011 counts per second. At each step, the fluorescence spectrum was integrated for 1 s before moving to the adjacent sample location. Fluorescence X-rays were detected with a Vortex energy-dispersive silicon drift quad (4-in-1) detector fitted with an aluminium foil attenuator to reduce high-intensity calcium fluorescence and increase sensitivity to trace elements. Initial spectral processing consisted of screening for a suite of 25 trace elements. Whereas only Sr, Mn and Ca concentrations exhibited consistent variation among and within fishes, other elements (e.g. Se, Zn) were observed in select taxa.

2.3. Sampling with laser ablation-inductively coupled plasma mass spectrometry

The LA–ICPMS was conducted at Baker Laboratory at SUNY–ESF. Elemental map collection (SXFM) was complemented by collection of transect data using a NewWave UP-193 nm solid-state laser (Electro Scientific Industries, Inc.) connected to a PerkinElmer Elan DRC-e inductively coupled plasma mass spectrometer. Ablated material was moved via a helium/nitrogen gas mixture from the laser into the ionization chamber of the mass spectrometer. Operating parameters for the laser were 70% power, 10 Hz, $35-100 \,\mu\text{m}$ spot size and $3 \,\mu\text{m}\,\text{s}^{-1}$ travel time. NIST-610 [72], USGS standard MACS-3 and an in-house pellet made from ground and pressed Freshwater Drum (*Aplodinotus grunniens*) otoliths [73] were used as standards. One-minute washouts of the mass spectrometer were performed between samples to remove residues from previous samples and to collect background counts. Drift corrections were performed as necessary for each sample session.

2.4. Data processing and analyses

Data reduction and processing for SXFM were completed using the program PYMCA [74] and the visualization software 'PRAXES,' developed at CHESS, to produce two-dimensional elemental maps and spatially explicit numerical output. Data were imported into Microsoft EXCEL, where elements were normalized against Ca (millimole element per mole Ca [5]). A Python script was used to produce final, coloured two-dimensional maps of element-to-Ca ratios. These maps were overlaid on photographs of corresponding otolith sections to compare otolith growth checks with changes in microchemistry and correlate chemical changes with life-history events.

We used elemental data at the otolith margin to infer water conditions at or near point of capture. To infer migration, elements needed to do more than simply rise and fall on a seasonal basis, as this could simply be attributed to annual variation. Therefore, we inferred migration by identifying elements that behaved differently during different life-history stages (i.e. high X : Ca for several years). A second criterion was to identify simultaneously alternating elements (i.e. high X : Ca coupled with low Y : Ca, and vice versa).

For sedentary fishes, we were particularly interested in Mn:Ca, as this has been shown to be a marker for hypoxia [22] and acidic waters [75], typical characteristics of the environments in which these fishes were caught. While there are currently no laboratory studies determining that environmental Mn incorporates into otoliths in proportion to the Mn dissolved in surrounding waters, several studies have compared otolith Mn in wild fish to known environmental conditions [73,76–78], and the biogeochemistry of manganese as a redox participant is well understood [79]. Limburg *et al.* [22] demonstrated otolith Mn's usefulness as an indicator of hypoxia and anoxia. Blackwater lakes and flooded forests in the Amazon are known to create acidified, hypoxic conditions [34] that promote the dissolution of particulate Mn. We used a threshold of 2.2×10^{-5} Mn : Ca as the marker of hypoxic waters

following Limburg *et al.* [22]. Chemical markers were expected to remain relatively constant (allowing for slight yearly fluctuations) assuming fishes were indeed sedentary.

To infer migration of juvenile floodplain migrants, Mn : Ca was also of interest. Given the life history of these fishes (spawning in white water, then dispersing into the floodplains and often black water), we expected to find low or undetectable Mn : Ca in the juvenile stage. When they migrate into black water to join adult fishes, we expected to see a marked rise in Mn : Ca. We used the same threshold of 2.2×10^{-5} Mn : Ca to infer entrance into black water.

To infer estuary use in migratory catfishes, we were particularly interested in the co-behaviour of Sr and Ba, as both of these elements have been used to identify migrations into and out of estuaries [80,81]. A recent study demonstrated the advantage of simultaneous use of Sr : Ca and Ba : Ca to identify migrations of European eels (*Anguilla anguilla*) from marine to estuarine to freshwater environments [82]. Following Tabouret *et al.* [82], migration into the estuary from the river channel was expected to be marked by a rise in Sr : Ca coupled with a drop in Ba : Ca.

Given the dearth of studies on Amazonian fish otolith chemistry, a broad suite of elements was selected to identify additional elements that may also be of interest but have rarely (or not at all) been reported in previous studies. New environmental markers were identified using the criteria previously outlined and comparisons with published environmental data.

A limitation of this study was the absence of water chemistry data available for all elements in Amazonian environments against which to compare otolith chemistry patterns. Campana [5] notes, however, that looking across multiple specimens in order to establish patterns is a robust technique even without environmental data.

3. Results

The main results of this study were: (i) otolith data from *Arapaima* sp. were consistent with a 'sedentary' life history involving no long-distance migrations, while the *Plagioscion* otolith bore markers indicating migration among chemically distinct environments; (ii) the majority of sampled *Prochilodus* bore markers consistent with migration from white water to black water; and (iii) *B. rousseauxii* otoliths from both sampled locations had chemical markers consistent with migration into and residency in the Amazon estuary, while *B. filamentosum* lacked these markers. We also found evidence of pollution exposure in two *Prochilodus* individuals as well as a possible new marker for Andean headwaters in *Brachyplatystoma* species.

3.1. Sedentary fishes

Our estimated age for the *Arapaima* sp. used in this study is 5 years. Sr : Ca in the *Arapaima* otolith showed repeated, annual banding, though a large area of discontinuity occurred near the sulcal groove (figure 1). By contrast, this Sr-depleted region contained elevated Zn, an element that appeared only in discrete zones in the otolith, suggesting differential crystallization (figure 1).

We estimated the age of the *P. squamosissimus* to be 15 years. SXFM analysis revealed alternating bands of Sr: Ca and Mn: Ca in this fish that occurred in distinct chemical 'zones' (figure 2): thick bands of high and low Sr: Ca alternating with one another every 3–4 years. Bands of elevated Mn: Ca appeared during periods of relatively low Sr: Ca, each zone representing several years. Thin bands of high Mn: Ca co-occurred with the outermost growth checks as well as on the marginal edge of the otolith.

3.2. Floodplain migrants

The ages of the *Prochilodus* were difficult to determine using otolith rings. Furthermore, the otoliths used in this study (lapilli) were different from those used in Silva & Stewart [48] (asterisci). Therefore, we divided fish into age classes using the length–frequency data from Silva [47] and Silva & Stewart [48]. We estimated that our samples included four age-0 fish (92–127 mm S.L.), one age-1 fish (197 mm S.L.) and one age-2 fish (267 mm S.L.).

LA–ICPMS data showed that otoliths of five out of six of our *P. nigricans* specimens recorded a rise, plateau and subsequent drop in Mn : Ca that began about 600–800 µm away from the otolith core (figure 3). Each rise in Mn : Ca was preceded by a visible growth check on the otolith (although not an annulus; figure 3). Two out of six of the *P. nigricans* otoliths also showed a considerable spike in Ba : Ca (figure 4).

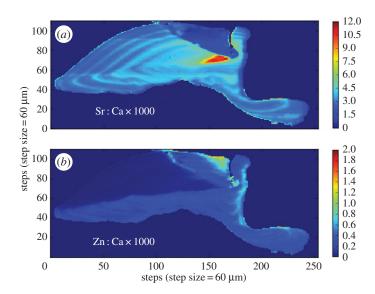


Figure 1. Otolith from Guyana-caught *Arapaima* sp. (*a*) Sr : Ca map; (*b*) Zn : Ca map.

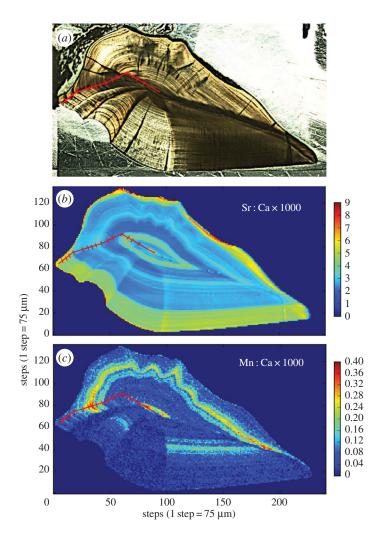


Figure 2. Otolith from Ecuadorian-caught *Plagioscion squamosissimus*. (*a*) Optical image (reflected light); (*b*) Sr : Ca map and (*c*) Mn : Ca map. The red transect marks putative annuli.

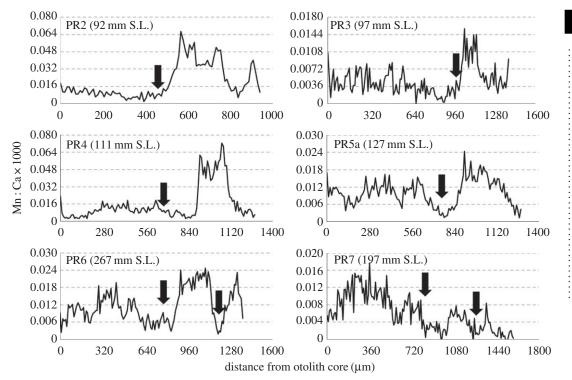


Figure 3. Plots show the variation in Mn : Ca over the lifetime of Ecuadorian-caught *Prochilodus nigricans* measured along a transect from the core of the otolith to its outer edge. Arrows mark the location of growth marks on the otolith. Note that we did not identify these marks as annuli. Different axes are used for each fish to emphasize the patterns of the data.

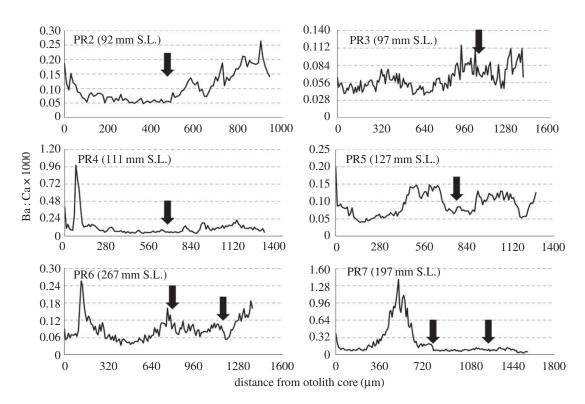


Figure 4. Plots show the variation in Ba : Ca over the lifetime of Ecuadorian-caught *Prochilodus nigricans* measured along a transect from the core of the otolith to its outer edge. Arrows mark the location of growth marks on the otolith. Note that we did not identify these marks as annuli. Different axes are used for each fish to emphasize the patterns of the data.

8

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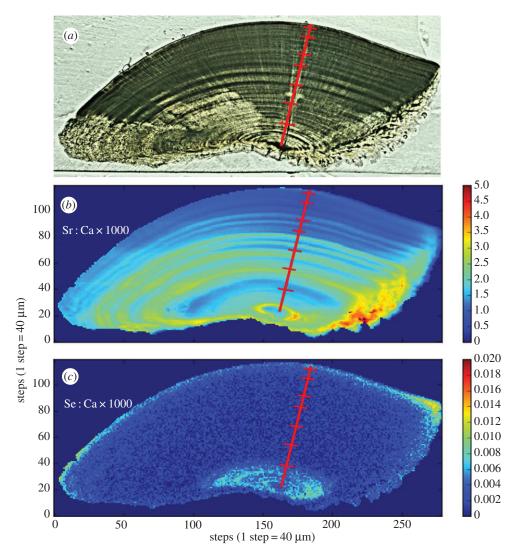


Figure 5. Otolith from Colombian-caught *Brachyplatystoma rousseauxii*. (*a*) Optical image (reflected light). (*b*) Sr : Ca map. (*c*) Se : Ca map. The red transect marks putative biannuli.

3.3. Long-distance migrants

We estimated the ages of the Colombian *B. rousseauxii* and *B. filamentosum* to be 3.5 and 4 years, respectively, and the Ecuadorian *B. rousseauxii* at 3 years following Alonso & Fabré [67]. SXFM revealed alternating bands of Se and Sr in *B. rousseauxii* and *B. filamentosum* that were delineated by biannual growth checks (figures 5 and 6). Marked Se deposition co-occurred with rapid growth in the early life of both *Brachyplatystoma* species, and repeated annually in *B. filamentosum* (figures 5 and 6). LA–ICPMS measurements in both *B. rousseauxii* otoliths showed a decoupling of Sr: Ca and Ba: Ca beginning 500–1000 µm away from the core, in which Sr:Ca rose and plateaued (with annual variation), while Ba: Ca declined and plateaued (figure 7). Around 3000 µm from the core, Sr: Ca declined and Ba: Ca rose and declined simultaneously (figure 7). Sr: Ca in both *B. rousseauxii* specimens indicated two distinct life-history phases: one marked by high, variable Sr: Ca during the first 1.5–2 years of life followed by a phase marked by lower, less variable Sr: Ca (figures 5 and 7).

4. Discussion

This study offers new insights into South American fish life history and contributes to the growing field of Amazonian fish otolith microchemistry. Using LA-ICPMS and SXFM in combination provided the

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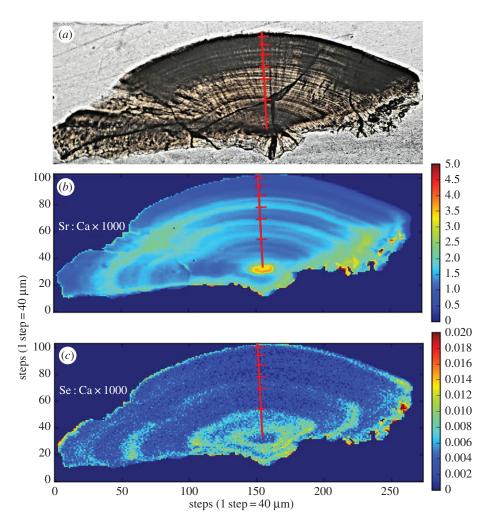
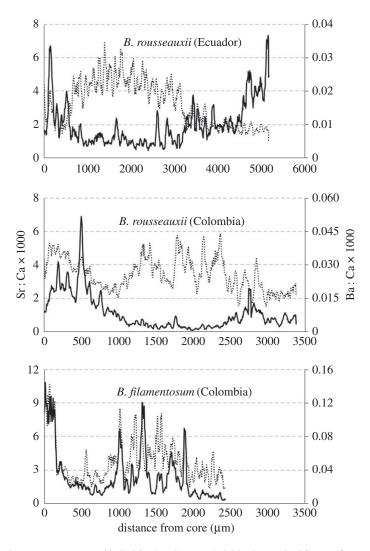


Figure 6. Otolith from Colombian-caught *Brachyplatystoma filamentosum*. (*a*) Optical image (reflected light). (*b*) Sr : Ca map. (*c*) Se : Ca map. The red transect marks putative biannuli.

flexibility to sample many elements simultaneously, which is particularly important in poorly studied systems where it is largely unknown which elements may be useful environmental markers and which may not. This study was not only simple and cost-effective, but also demonstrated how powerful these analytical techniques can be for studying imperilled systems. With a limited sample size, we identified potential new markers for critical habitats and elucidated the life-history complexity of several fish species, while avoiding the issues associated with tagging–tracking methods often used in life-history studies. This study lays the groundwork for a potentially very productive line of research that could do much to provide the necessary biological and ecological basis for conserving these important commercial fishes.

Our results for *Arapaima* sp. were consistent with previous findings that this fish does not migrate among chemically distinct environments. Castello [39] demonstrated that *Arapaima* in the central Amazon appear to migrate short distances among eight different habitats over the course of a flood cycle as water levels regulate habitat availability. However, these habitats are in relatively close proximity with one another and are expected to have roughly the same chemistry. The Sr:Ca observed in the present study (figure 1) oscillated minimally on an annual basis, which would easily be explained by the seasonal introduction of flood waters into the lake from which this fish was captured. The detected Zn (figure 1), which was unique among the fishes in this study, also fits the known ecology of *Arapaima*, outlined above. Zn has been shown to vary inversely with oxygen concentration and pH in the Orinoco River system [83]. The conditions associated with elevated Zn concentrations are common in environments like the lakes that *Arapaima* inhabit. The lifetime Zn:Ca patterns observed in this fish were similar to those of Sr:Ca, displaying small oscillations that could also be explained by annual inundations of the local environment. However, it is also possible that Zn:Ca is under



11

Figure 7. Plots show the variation in Sr : Ca (dashed line) and Ba : Ca (solid line) over the lifetime of two *B. rousseauxii* and one *B. filamentosum*, measured along a transect from the core of the otolith to its outer edge. Different axes are used for each fish to emphasize the patterns of the data. In each graph, the elemental ratio transects were smoothed with 5-point moving averages.

physiological control. Limburg & Elfman [84] found that Zn:Ca in the otoliths of salmoniform and esociform fishes varied seasonally and correlated positively with growth rate, suggesting an underlying physiological mechanism. Additional studies are required to elucidate this in *Arapaima*. Regardless, the otolith microchemistry of this individual appears to be consistent with what one would expect from a typical sedentary fish that undertakes only short-distance lateral migrations among chemically similar environments.

The distinct chemical 'zones' of high/low Sr:Ca and high/low Mn:Ca (figure 2) found in the *Plagioscion* otolith suggest that this individual spent multiple growing seasons in several chemically distinct environments. This fish was caught in the lower reaches of the blackwater Cuyabeno drainage near its confluence with the whitewater Rio Aguarico, both of which are surrounded by many floodplain lakes. Therefore, it seems reasonable to conclude that these environments may have been among those used by this fish. Potential migration among such diverse environments contrasts with the behaviour of *Arapaima* sp., whose otolith microchemistry was best explained by short-distance migrations among chemically similar environments. These results suggest that *Plagioscion* may have a more complex life history than previously thought and, furthermore, may be behaviourally distinct from traditional sedentary fishes (e.g. *Arapaima*). Further studies with more extensive sampling will be necessary to better clarify these life-history details and determine how common these hypothesized behaviours may be within this widespread species.

Mn: Ca was also shown to be potentially useful as a marker for black water, which is a critical environment in the life histories of *Plagioscion* and *Prochilodus*. The Mn: Ca values of the thick Mn: Ca

bands found in the *Plagioscion* otolith exceeded the hypoxia threshold of 2.2×10^{-5} used in this study [22], indicating a water chemistry consistent with preferred spawning conditions (i.e. low dissolved oxygen) identified by Baumgartner *et al.* [42] and Bialetzki *et al.* [43]. The age-0 *Prochilodus* used in this study were probably born in April 1999 during the spawning season and caught for use by Silva & Stewart [48] in September 1999 shortly after having migrated upstream. The otolith growth checks preceding elevated Mn : Ca may mark this migration into black water, where these fishes were caught. Therefore, given that *Prochilodus* spawns in white water, and we found no Mn in strictly whitewater fishes (i.e. both *Brachyplatystoma* species), we hypothesize that the first Mn : Ca peak observed in *Prochilodus* marks the first migration from downstream whitewater nursery habitats to the upstream blackwater rivers.

Sr: Ca and Ba: Ca data suggest that the B. rousseauxii individuals used in this study lived their first 1.5-2 years in the Amazon estuary. Using only Sr: Ca, two life-history stages are clearly visible (i.e. the first 1.5–2 years and then thereafter) in *B. rousseauxii* (figures 5b and 7). Sr: Ca in otoliths has been demonstrated to increase proportionally with salinity, reflecting the higher concentrations of Sr relative to Ca found in marine waters [80,85,86], and has been used in many studies as a marker in anadromous fishes to indicate a shift between freshwater and marine environments [87,88], including in other tropical species [89,90]. Furthermore, the Sr concentration found in the Amazonian endmember and its tributaries has been shown to be an order of magnitude lower than that found in marine waters [91,92]. The variation in Sr: Ca that we observed is comparable with the differences seen in studies of diadromous fishes that migrate between freshwater and marine environments (e.g. [82,85,88,93,94]). The observed Ba: Ca patterns were also consistent with the migration of a fish from freshwater into an estuary. Elsdon & Gillanders [81] found that otoliths from black bream *Acanthopagrus butcheri* migrating between freshwater rivers and saltwater estuaries had roughly double the otolith Ba: Ca while in freshwater versus while in the estuary, which reflected ambient water chemistry. Recent studies have also shown that Sr and Ba can be used in conjunction to demonstrate the transition between freshwater rivers and estuaries [82,95,96]. For example, the mirrored behaviours of otolith Ba: Ca and Sr: Ca in B. rousseauxii (figures 5 and 7) were nearly identical to those observed in European eels [82] and barramundi Lates calcarifer [95,96] migrating from freshwater into an estuary. Prior studies using this technique have identified estuary use by fish species with distinctly marine life-history stages, such as hilsa Tenualosa ilisha (e.g. [89]) and barramundi (e.g. [96]). However, Brachyplatystoma species are thought to remain in freshwater parts of the estuary [27]. If verified by additional studies, these results would mark the first time that this technique has been used to identify estuary use in a tropical migratory fish that lacks a defined marine life-history stage. Therefore, this technique would stand as a powerful tool for identifying residence in this critically important environment.

While the *B. rousseauxii* results were consistent with those of fishes migrating into an estuary from a river, our data suggested that *B. filamentosum* did not migrate passively or actively into the estuary. Rather, Sr:Ca and Ba:Ca for *B. filamentosum* rose and declined simultaneously (figure 7). These results are consistent with those of Hegg *et al.* [17], suggesting that some *Brachyplatystoma* species use the estuary as a nursery area, while others may never reach the estuary and therefore have different life-history strategies.

Our data also indicate that Se may be a useful marker for identifying migration into or out of Andean headwaters. There are few published studies on Se in otoliths and most use it to identify severely polluted environments (e.g. [13,97,98]). However, Yee *et al.* [99] measured Se concentrations in tributaries of the Orinoco Basin, finding that concentrations were highest in tributaries originating in the Andes Mountains (white water), low in the main channel and very low in other tributaries such as those draining the Guyana shield. Because the whitewater tributaries of the Amazon also originate in the Andes, the Amazon and Orinoco Basins probably have a similar pattern of Se concentrations among tributaries. Se:Ca in the *B. rousseauxii* from Colombia was high during roughly the first few months of life, but zero during its assumed residence in the estuary, up until when it was caught in the Rio Caqueta (an Andean tributary), where we found Se:Ca again at the otolith margin (figure 6). The repeated and alternating banding of high Sr:Ca/Ba:Ca and Se:Ca therefore suggest that the Colombian *B. filamentosum* in this study may have migrated into and out of the same or similar headwater tributaries annually during its lifetime (figures 6 and 7), but did not establish residence in those headwater areas.

We hypothesize that the very high Ba : Ca peaks present in two *Prochilodus* specimens were the result of environmental contamination. These values were an order of magnitude higher than those observed anywhere else in the otolith (figure 4) and were the highest concentrations ever observed by one of us (K.E.L.). Both left and right otoliths were analysed on one individual to verify this result. Given that these fish were caught in the Ecuadorian Amazon just downstream from the Lago Agrio oil field, pollution seems a possible cause. Ecuador has a history of poor oil field management since oil exploitation began in 1972, leading to environmental and public health concerns such as increased cancer risk and birth defects among people living nearby [62,100-104]. Measured Ba concentrations from drilling wells at Lago Agrio were as high as $10\,100 \text{ mg kg}^{-1}$ due to barite contamination; even so-called remediated wells had concentrations over 1000 mg kg^{-1} [105]. Barite (BaSO₄) is a common additive to 'drilling mud', the lubricant used in oil drilling processes [106], and is known to be bioavailable in marine systems, particularly to detritivores [107]. Although largely insoluble near neutral pH, barite acts as a source of reducible sulfate for sulfate-reducing bacteria [108,109], a process that releases Ba into pore waters [110]. Neff [107] also suggested that a fish's stomach may provide a sufficiently acidic environment for dissolution. Further study of the fishes in this region is needed to better describe this phenomenon.

5. Conclusion

The data and inferences on the life histories of five Neotropical freshwater fishes reported here provide a foundation upon which to develop testable hypotheses, thereby setting priorities and directions for future studies. Extensive water sampling, both spatially and temporally, will be critical to accurately verify and recreate the migratory strategies hypothesized herein. Ideal future studies should include water sampling at a scale relevant to the species in question (i.e. basin-wide for *Brachyplatystoma*, or regionally for *Prochilodus*). Furthermore, a comparative, basin-wide approach to fish sampling should be taken to elucidate differences among distinct populations (e.g. Ecuadorian versus central Amazonian *Prochilodus*) and/or species (e.g. *Brachyplatystoma* species). Finally, the mechanisms by which newly identified chemical markers were incorporated into otoliths need to be better understood via laboratory experiments and comparison of Amazonian water chemistry to otolith chemistry from fishes caught in those same waters. As demonstrated by this study, a combination of LA–ICPMS and SXFM can facilitate unravelling the complex life histories of these fishes. Subsequent larger studies should provide the migration and habitat use data that are fundamental to developing sound management and conservation strategies for these species. There is also considerable potential for applications of these methods to fishes in other large, tropical rivers of the world that have similar emerging threats [111].

Ethics. Permission to collect samples of *Arapaima* in Guyana was granted to LC Watson and DJ Stewart by the Guyana Environmental Protection Agency (permit no. 060706 BR 055), and the study was reviewed and approved by the Institutional Animal Care and Use Committee (Protocol No. 06-02) of SUNY College of Environmental Science and Forestry. EA Silva, an Ecuadorian citizen working on a thesis research project in 1999 in the Ecuadorian Amazon, collected otoliths of *Prochilodus*; he also obtained otoliths of *Brachyplatystoma* and *Plagioscion* from commercially harvested fishes. No special collecting permit or 'Animal Care Protocol' was required at that time. Otoliths of *Brachyplatystoma* from the Caquetá River in Colombia were all taken by DL Munoz-Sosa in 1995–1996 from commercially harvested fishes. No special collecting permit or 'Animal Care Protocol' was required at the time; he was a Colombian citizen working on a thesis research project. Field efforts in Colombia were supported by Instituto de Investigaciones Amazonicas—SINCHI and the Fulbright Commission.

Data accessibility. Our data are deposited at Dryad: http://dx.doi.org/10.5061/dryad.8kd35.

Authors' contributions. T.W.H., D.J.S. and K.E.L. designed the study. T.W.H. prepared all samples for analysis. T.W.H. and K.E.L. collected and analysed the data. T.W.H., D.J.S., K.E.L. and L.C. interpreted the results and wrote the manuscript. All authors gave final approval for publication.

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References

 Mugiya Y, Watabe N, Yamada J, Dean J, Dunkelberger D, Shimizu M. 1981 Diurnal rhythm in otolith formation in the goldfish, *Carassius auratus. Comp. Biochem. Physiol. A. Physiol.* **68**, 659–662. (doi:10.1016/0300-9629(81) 90374-1)

 Pannella G. 1971 Fish otoliths: daily growth layers and periodical patterns. *Science* 173, 1124–1127. (doi:10.1126/science.173.4002.1124)
 Campana SE, Neilson JD. 1985 Microstructure of fish otoliths. *Can. J. Fish. Aquat. Sci.* 42, 1014–1032. (doi:10.1139/f85-127)

- Elsdon TS, Wells BK, Campana SE, Gillanders BM, Jones CM, Limburg KE, Secor DH, Thorrold SR, Walther BD. 2008 Otolith chemistry to describe movements and life-history parameters of fishes: hypotheses, assumptions, limitations and inferences. Oceanogr. Mar. Biol. Annu. Rev. 46, 297-330. (doi:10.1201/9781420065756.Ch7)
- 5 Campana SE. 1999 Chemistry and composition of fish otoliths: pathways, mechanisms and applications. Mar. Ecol. Prog. Ser. 188, 263–297. (doi:10.3354/meps188263)
- 6. Campana SE, Thorrold SR. 2001 Otoliths, increments, and elements: keys to a comprehensive understanding of fish populations? Can. J. Fish. Aquat. Sci. 58, 30-38. (doi:10.1139/ f00-177)
- Wells BK, Rieman BE, Clayton JL, Horan DL, Jones 7. CM. 2003 Relationships between water, otolith, and scale chemistries of westslope cutthroat trout from the Coeur d'Alene River, Idaho: the potential application of hard-part chemistry to describe movements in freshwater. Trans. Am. Fish. Soc. 132, 409-424. (doi:10.1577/1548-8659(2003)132 < 0409: RBW0AS>2.0.C0;2)
- Muhlfeld CC, Marotz B, Thorrold SR, FitzGerald JL. 8. 2005 Geochemical signatures in scales record stream of origin in westslope cutthroat trout. Trans. Am. Fish. Soc. 134, 945-959. (doi:10.1577/T04-029.1)
- 9. Crook DA, Gillanders BM. 2006 Use of otolith chemical signatures to estimate carp recruitment sources in the mid-Murray River, Australia, River Res. Appl. 22, 871-879. (doi:10.1002/rra.941)
- Clarke AD, Telmer KH, Shrimpton J. 2007 Habitat 10. use and movement patterns for a fluvial species, the Arctic grayling, in a watershed impacted by a large reservoir: evidence from otolith microchemistry. J. Appl. Ecol. 44, 1156-1165. (doi:10.1111/j.1365-2664.2007.01350.x)
- 11 Campana S, Pangle KL, Ludsin SA, Fryer BJ. 2010 Otolith microchemistry as a stock identification tool for freshwater fishes: testing its limits in Lake Erie. Can. J. Fish. Aquat. Sci. 67, 1475-1489. (doi:10.1139/F10-076)
- 12. Muhlfeld CC, Thorrold SR, McMahon TE, Marotz B, Gillanders B. 2012 Estimating westslope cutthroat trout (Oncorhynchus clarkii lewisi) movements in a river network using strontium isoscapes. Can. J. Fish. Aquat. Sci. 69, 906-915. (doi:10.1139/ f2012-033)
- 13. Limburg KE, Lochet A, Driscoll D, Dale DS, Huang R. 2010 Selenium detected in fish otoliths: a novel tracer for a polluted lake? Environ Biol Fishes 89. 433-440. (doi:10.1007/s10641-010-9671-4)
- Halden N, Friedrich L. 2008 Trace-element 14. distributions in fish otoliths: natural markers of life histories, environmental conditions and exposure to tailings effluence. Mineral Mag. 72, 593–605. (doi:10.1180/minmag.2008.072.2.593)
- Pracheil BM, Hogan JD, Lyons J, McIntyre PB. 2014 15. Using hard-part microchemistry to advance conservation and management of North American freshwater fishes. Fisheries 39, 451-465. (doi:10.1080/03632415.2014. 937858)
- 16. Barthem R, Goulding M. 2007 An unexpected ecosystem: the Amazon as revealed by fisheries. Lima, Peru: Amazon Conservation Association: Missouri Botanical Garden Press.

- 17. Hegg JC, Giarrizzo T, Kennedy BP. 2015 Diverse early life-history strategies in migratory Amazonian catfish: implications for conservation and management. PLoS ONE 10, e0129697. (doi:10.1371/journal.pone.0129697)
- Garcez R, Humston R, Harbor D, Freitas CE. 2014 18 Otolith geochemistry in young-of-the-year peacock bass Cichla temensis for investigating natal dispersal in the Rio Negro (Amazon-Brazil) river system. Ecol. Freshw. Fish. 24, 242-251. (doi:10.1111/eff.12142)
- 19 Pouilly M, Point D, Sondag F, Henry M, Santos RV. 2014 Geographical origin of Amazonian freshwater fishes fingerprinted by 87Sr/86Sr ratios on fish otoliths and scales. Environ. Sci. Technol. 48, 8980-8987. (doi:10.1021/es500071w)
- Sousa R, Humston R, Freitas C. 2016 Movement 20. patterns of adult peacock bass Cichla temensis between tributaries of the middle Negro River basin (Amazonas-Brazil): an otolith geochemical analysis. Fish. Manage. Ecol. 23, 76-87. (doi:10.1111/fme.12166)
- 21. Duponchelle F et al. In press. Trans-Amazonian natal homing in giant catfish. J. Appl. Ecol. (doi:10.1111/1365-2664.12665)
- 22. Limburg KE, Walther BD, Lu Z, Jackman G, Mohan J, Walther Y, Nissling A, Weber PK, Schmit AK. 2015 In search of the dead zone: Use of otoliths for tracking fish exposure to hypoxia. J. Mar. Syst. 141, 167-178. (doi:10.1016/j.jmarsys.2014.02.014)
- 23. Limburg KE, Høie H, Dale DS. 2010 Bromine patterns in Norwegian coastal Cod otoliths-a possible marker for distinguishing stocks? Environ. Biol. Fishes 89, 427-432. (doi:10.1007/s10641-010-9660-7)
- 24. Limburg KE, Huang R, Bilderback DH. 2007 Fish otolith trace element maps: new approaches with synchrotron microbeam X-ray fluorescence. X-Ray Spectrom. 36, 336-342. (doi:10.1002/xrs.980)
- 25. Bayley PB, Petrere Jr M. 1989 Amazon fisheries: assessment methods, current status and management options. In Proc. Int. Large River Symp. (ed. DP Dodge), pp. 385-398. Ottawa, CA: Canadian Special Publication of Fisheries and Aquatic Sciences.
- 26. Godinho AL, Kynard B, Godinho HP. 2007 Migration and spawning of female surubim (Pseudoplatystoma corruscans, Pimelodidae) in the São Francisco river, Brazil, Environ, Biol, Fishes 80. 421-433. (doi:10.1007/s10641-006-9141-1)
- Barthem R, Goulding M. 1997 The catfish 27. connection: ecology, migration, and conservation of Amazon predators. New York, NY: Columbia University Press.
- 28. Gibbs RJ. 1967 The geochemistry of the Amazon River system: part I. The factors that control the salinity and the composition and concentration of the suspended solids. Geol. Soc. Am. Bull. 78, 1203-1232. (doi:10.1130/0016-7606(1967)78 [1203:TGOTAR12.0.CO:2)
- 29. Irion G, Junk WJ, Mello JASN. 1997 The large central Amazonian river floodplains near Manaus: geological, climatological, hydrological and geomorphological aspects. In The central Amazon floodplain: ecology of a pulsing system (ed. WJ Junk), pp. 23-46. Berlin, Germany: Springer.
- 30. Lowe-McConnell RH. 1987 Ecological studies in tropical fish communities. Cambridge, UK: Cambridge University Press.

- 31. Junk WJ, Bayley PB, Sparks RE. 1989 The flood pulse concept in river floodplain systems. In Proc. Int. Large River Symp. (ed. DP Dodge), pp. 110-127. Ottawa, CA: Canadian Special Publication of Fisheries and Aquatic Sciences.
- 32. Goulding M. 1980 The fishes and the forest: explorations in Amazonian natural history. Berkeley, CA: University of California Press.
- Winemiller KO, Jepsen D. 1998 Effects of 33. seasonality and fish movement on tropical river food webs. J. Fish Biol. 53, 267-296. (doi:10.1111/j.1095-8649.1998.tb01032.x)
- Petry P, Bayley P, Markle D. 2003 Relationships 34. between fish assemblages, macrophytes and environmental gradients in the Amazon River floodplain. J. Fish Biol. 63, 547-579. (doi:10.1046/j.1095-8649.2003.00169.x)
- 35. Junk W, Soares G, Carvalho F. 1983 Distribution of fish species in a lake of the Amazon river floodplain near Manaus (Lago Camaleão), with special reference to extreme oxygen conditions. Amazoniana 7, 397-431.
- 36. Camargo M, Isaac VJ, Fernandes MEB. 2003 Ictiofauna estuarina. In Os manguezais da costa Norte do Brasil. (ed. MEB Fernandes), pp. 105–142.São Luís, Brasil: Fundação Rio Bacanga.
- 37. Barthem RB, de Brito Ribeiro MCL, Petrere MJr. 1991 Life strategies of some long-distance migratory catfish in relation to hydroelectric dams in the Amazon Basin. Biol. Conserv. 55, 339-345. (doi:10.1016/0006-3207(91)90037-A)
- 38. Schwassmann H. Barthem R. Carvalho M. 1989 A note on the seasonally shifting zone of high primary production in the Bay of Marajó, Pará, Brazil, 1983–1984. Acta Bot. Bras. 2, 165–170. (doi:10.1590/S0102-33061988000300014)
- 39. Castello L. 2008 Lateral migration of Arapaima gigas in floodplains of the Amazon. Ecol. Freshw. Fish 17, 38-46.
 - (doi:10.1111/j.1600-0633.2007.00255.x)
- 40. Loubens G, Panfili J. 2003 Biologie de Plagioscion squamosissimus (Teleostei: Sciaenidae) dans le bassin du Mamoré (Amazonie bolivienne). Ichthyol. Explor. Freshw. 14, 335-352.
- 41. Daga VS, Gogola TM, Sanches PV, Baumgartner G, Baumgartner D, Piana PA, Gubiani ÉA, Delariva RL. 2009 Fish larvae assemblages in two floodplain lakes with different degrees of connection to the Paraná River, Brazil, Neotrop, Ichthyol, 7, 429-438. (doi:10.1590/S1679-62252009000300010)
- Baumgartner M, Nakatini K, Baumgartner G, 42. Makrakis M. 2003 Spatial and temporal distribution of 'curvina' larvae (Plagioscion squamosissimus Heckel, 1840) and its relationship to some environmental variables in the upper Paraná River floodplain, Brazil. Braz. J. Biol. 63, 381-391. (doi:10.1590/S1519-69842003000300004)
- 43. Bialetzki A, Nakatani K, Sanches PV, Baumgartner G. 2004 Eggs and larvae of the 'curvina' Plagioscion squamosissimus (Heckel, 1840) (Osteichthyes, Sciaenidae) in the Baía River, Mato Grosso do Sul State, Brazil. J. Plankton Res. 26, 1327-1336. (doi:10.1093/plankt/fbh123)
- 44. Borie A, Mok H, Chao N, Fine M. 2014 Spatiotemporal variability and sound characterization in silver croaker Plagioscion squamosissimus (Sciaenidae) in the Central Amazon, PLoS ONE 9, e99326. (doi:10.1371/journal.pone.0099326)

- Barbosa N, Rocha R, Lucena Frédou F. 2012 The reproductive biology of *Plagioscion* squamosissimus (Heckel, 1840) in the Pará River estuary (Amazon Estuary). J. Appl. Ichthyol. 28, 800–805. (doi:10.1111/j.1439-0426.2012.02040.x)
- Araujo-Lima CARM, Ruffino ML. 2003 Migratory fishes of the Brazilian Amazon. In *Migratory fishes* of South America (eds J Carolsfeld, B Harvey, C Ross, A Baer), pp. 233–301. Victoria, Canada: IDRC and The World Bank.
- Silva EA. 2000 Life history and migration patterns of the commercial fish *Prochilodus nigricans* (bocachico) in northeastern Ecuador. MS Thesis. Syracuse, State University of New York, College of Environmental Science and Forestry, Syracuse, NY, USA.
- Silva EA, Stewart DJ. 2006 Age structure, growth and survival rates of the commercial fish *Prochilodus nigricans* (bocachico) in North-eastern Ecuador. *Environ. Biol. Fishes* 77, 63–77. (doi:10.1007/s10641-006-9055-y)
- Batista JS, Alves-Gomes JA. 2006 Phylogeography of *Brachyplatystoma rousseauxii* (Siluriformes -Pimelodidae) in the Amazon Basin offers preliminary evidence for the first case of 'homing' for an Amazonian migratory catfish. *Gen. Mol. Res.* 5, 723–740.
- Cañas C, Pine W. 2011 Documentation of the temporal and spatial patterns of pimelodidae catfish spawning and larvae dispersion in the Madre de Dios River (Peru): Insights for conservation in the Andean Amazon headwaters. *River Res. Appl.* 27, 602–611. (doi:10.1002/rra. 1377)
- Castello L, McGrath DG, Beck PS. 2011 Resource sustainability in small-scale fisheries in the Lower Amazon floodplains. *Fish. Res.* **110**, 356–364. (doi:10.1016/j.fishres.2011.05.002)
- Coe MT, Costa MH, Soares-Filho BS. 2009 The influence of historical and potential future deforestation on the stream flow of the Amazon River–land surface processes and atmospheric feedbacks. J. Hydrol. 369, 165–174. (doi:10.1016/j.jhydrol.2009.02.043)
- Castello L, McGrath DG, Hess LL, Coe MT, Lefebvre PA, Petry P, Macedo MN, Renó VF, Arantes CC. 2013 The vulnerability of Amazon freshwater ecosystems. *Conserv. Lett.* 6, 217–229. (doi:10.1111/conl.12008)
- Petrere M, Barthem RB, Cordoba EA, Gomez BC.
 2004 Review of the large catfish fisheries in the upper Amazon and the stock depletion of piraiba (*Brachyplatystoma filamentosum* Lichtenstein). *Rev. Fish Biol. Fish.* 14, 403–414. (doi:10.1007/s11160-004-8362-7)
- Agostinho AA, Gomes LC, Fernandez DR, Suzuki HI. 2002 Efficiency of fish ladders for neotropical ichthyofauna. *River Res. Appl.* 18, 299–306. (doi:10.1002/tra.674)
- Fernandez DR, Agostinho AA, Bini LM, Pelicice FM. 2007 Diel variation in the ascent of fishes up an experimental fish ladder at Itaipu Reservoir: fish size, reproductive stage and taxonomic group influences. *Neatrop. Ichthyol.* 5, 215–222. (doi:10.1590/S1679-62252007000200016)
- Makrakis S, Makrakis MC, Wagner RL, Dias JHP, Gomes LC. 2007 Utilization of the fish ladder at the Engenheiro Sergio Motta Dam, Brazil, by long distance migrating potamodromous species.

Neotrop. lchthyol. 5, 197–204. (doi:10.1590/S1679-62252007000200014)

- Oldani NO, Baguin CRM, Nestler JM, Goodwin RA. 2007 Is fish passage technology saving fish resources in the lower La Plata River basin? *Neotrop. Ichthyol.* 5, 89–102. (doi:10.1590/S1679-62252007000200002)
- Ashe K. 2012 Elevated mercury concentrations in humans of Madre de Dios, Peru. PLoS ONE 7, e33305. (doi:10.1371/journal.pone.0033305)
- Neill C, Deegan LA, Thomas SM, Cerri CC. 2001 Deforestation for pasture alters nitrogen and phosphorus in small Amazonian streams. *Ecol. Appl.* **11**, 1817–1828. (doi:10.1890/1051-0761 (2001)011[1817:DFPANA]2.0.C0;2)
- Finer M, Jenkins CN, Pimm SL, Keane B, Ross C. 2008 Oil and gas projects in the western Amazon: threats to wilderness, biodiversity, and indigenous peoples. *PLoS ONE* 8, e2932. (doi:10.1371/journal.pone.0002932)
- Sebastián MS, Hurtig AK. 2004 Oil exploitation in the Amazon basin of Ecuador: a public health emergency. *Rev. Panam. Salud Públ.* 15, 205–211. (doi:10.1590/s1020-49892004000300014)
- Ellgehausen H, Guth JA, Esser HO. 1980 Factors determining the bioaccumulation potential of pesticides in the individual compartments of aquatic food chains. *Ecotoxicol. Environ. Saf.* 4, 134–157. (doi:10.1016/0147-6513(80)90015-9)
- Mergler D, Anderson HA, Chan LHM, Mahaffey KR, Murray M, Sakamoto M, Stern AH. 2007 Methylmercury exposure and health effects in humans: a worldwide concern. *Ambio* 36, 3–11. (doi:10.1579/0044-7447(2007)36[3:MEAHEI]2.0.C0;2)
- Secor DH, Dean JM, Laban EH. 1991 Manual for otolith removal and preparation for microstructural examination. Columbia, SC: Belle W. Baruch Institute, University of South Carolina Press.
- 66. Watson LC. 2011 Ecology and conservation of *Arapaima* in Guyana: recovery of a giant fish in distress. Doctoral dissertation. State University of New York, College of Environmental Science and Forestry, Syracuse, NY, USA.
- 67. Alonso JC, Fabré NN. 2003 Spatial and temporal pattern of the population structure and current state of fishing exploitation of the dourada (*Brachyplatystoma flavicans*, Lichtenstein, 1819) along the system estuary Amazonas–Solimoes. In Second Int.Symp.on the Management of Large Rivers for Fisheries: Sustaining Livelihoods and Biodiversity in the New Millennium (eds RL Welcomme, T Petr). Phnom Penh, Cambodia: FAO & Mekong River Commission.
- Arantes C, Castello L, Stewart D, Cetra M, Queiroz H. 2010 Population density, growth and reproduction of *Arapaima* in an Amazonian river-floodplain. *Ecol. Freshw. Fish.* 19, 455–465. (doi:10.1111/j.1600-0633.2010.00431.x)
- Barthem RB, Fabré NN. 2004 Biologia e diversidade dos recursos pesqueiros da Amazônia. In A pesca e os recursos pesqueiros na Amazônia brasileira. (ed. M Ruffino), pp. 17–62. Manaus, Brazil: ProVárzea.
- Bilderback D, Huang R, Kazimiro A, Kriksunov I, Limburg K, Fontes E. 2003 Monocapillary optics developments and applications. *Adv. X-Ray Anal.* 46, 320–325.
- Cornaby SW. 2008 The handbook of X-ray single-bounce monocapillary optics, including optical design and synchrotron applications.

Doctoral dissertation. Cornell University, Ithaca, NY, USA.

- NIST. Material Measurement Laboratory—Standard Reference Materials. 2015. See https://www-s.nist.gov/srmors/view_detail. cfm?srm=612
- Limburg KE, Olson C, Walther Y, Dale D, Slomp CP, Høie H. 2011 Tracking Baltic hypoxia and cod migration over millennia with natural tags. *Proc. Natl Acad. Sci. USA* 108, E177–E182.

(doi:10.1073/pnas.1100684108)

- Solé V, Papillon E, Cotte M, Walter P, Susini J. 2007 A multiplatform code for the analysis of energy-dispersive X-ray fluorescence spectra. *Spectrochim. Acta B* 62, 63–68. (doi:10.1016/j.sab.2006.12.002)
- Moreau G, Barbeau C, Frenette J, Saint-Onge J, Simoneau M. 1983 Zinc, manganese, and strontium in opercula and scales of brook trout (*Salvelinus fontinalis*) as indicators of lake acidification. *Can. J. Fish. Aquat. Sci.* 40, 1685–1691. (doi:10.1139/f83-195)
- Dorval E, Jones CM, Hannigan R, Montfrans JV. 2007 Relating otolith chemistry to surface water chemistry in a coastal plain estuary. *Can. J. Fish. Aquat. Sci.* 64, 411–424. (doi:10.1139/f07-015)
- Forrester GE. 2005 A field experiment testing for correspondence between trace elements in otoliths and the environment and for evidence of adaptation to prior habitats. *Estuaries* 28, 974–981. (doi:10.1007/BF02696025)
- Mohan JA, Rulifson RA, Corbett DR, Halden NM. 2012 Validation of oligohaline elemental otolith signatures of striped bass by use of in situ caging experiments and water chemistry. *Mar. Coast. Fish.* 4, 57–70. (doi:10.1080/19425120.2012.656533)
- Reddy KR, DeLaune RD. 2008 Biogeochemistry of wetlands: science and applications. Boca Raton, FL: CRC Press.
- Kraus RT, Secor DH. 2004 Incorporation of strontium into otoliths of an estuarine fish. *J. Exp. Mar. Biol. Ecol.* **302**, 85–106. (doi:10.1016/j.jembe.2003.10.004)
- Elsdon TS, Gillanders BM. 2005 Alternative life-history patterns of estuarine fish: barium in otoliths elucidates freshwater residency. *Can. J. Fish. Aquat. Sci.* 62, 1143–1152. (doi:10.1139/f05-029)
- Tabouret H, Bareille G, Claverie F, Pécheyran C, Prouzet P, Donard O. 2010 Simultaneous use of strontium:calcium and barium:calcium ratios in otoliths as markers of habitat: application to the European eel (*Anguilla anguilla*) in the Adour basin, South West France. *Mar. Environ. Res.* 70, 35–45. (doi:10.1016/j.marenvres.2010.02.006)
- Mora A, Baquero JC, Alfonso JA, Pisapia D, Balza L. 2010 The Apure River: geochemistry of major and selected trace elements in an Orinoco River tributary coming from the Andes, Venezuela. *Hydrol. Process.* 24, 3798–3810. (doi:10.1002/hyp.7801)
- Limburg KE, Elfman M. 2010 Patterns and magnitude of Zn:Ca in otoliths support the recent phylogenetic typology of Salmoniformes and their sister groups. *Can. J. Fish. Aquat. Sci.* 67, 597–604. (doi:10.1139/F10-014)
- Secor DH, Henderson-Arzapalo A, Piccoli P. 1995 Can otolith microchemistry chart patterns of migration and habitat utilization in anadromous

fishes? J. Exp. Mar. Biol. Ecol. 192, 15-33. (doi:10.1016/0022-0981(95)00054-U)

- 86. Thorrol SR, Jones CM, Campana SE. 1997 Response of otolith microchemistry to environmental variations experienced by larval and juvenile Atlantic croaker (Micropogonias undulatus). Limnol. Oceanogr. 42, 102-111. (doi:10.4319/lo.1997.42.1.0102)
- Brown RJ, Severin KP. 2009 Otolith chemistry 87. analyses indicate that water Sr:Ca is the primary factor influencing otolith Sr:Ca for freshwater and diadromous fish but not for marine fish. Can. J. Fish. Aquat. Sci. 66, 1790-1808. (doi:10.1139/F09-112)
- 88. Walther B, Limburg K. 2012 The use of otolith chemistry to characterize diadromous migrations. J. Fish Biol. 81, 796-825. (doi:10.1111/j.1095-8649.2012.03371.x)
- 89. Milton DA, Chenery SR. 2003 Movement patterns of the tropical shad hilsa (Tenualosa ilisha) inferred from transects of 87Sr/86Sr isotope ratios in their otoliths. Can. J. Fish. Aquat. Sci. 60, 1376-1385. (doi:10.1139/f03-133)
- 90. Milton DA, Chenery SR. 2005 Movement patterns of barramundi Lates calcarifer, inferred from 87Sr/86Sr and Sr/Ca ratios in otoliths, indicate non-participation in spawning. Mar. Ecol. Prog. Ser. 301, 279-291. (doi:10.3354/meps301279)
- 91. Palmer M, Edmond J. 1992 Controls over the strontium isotope composition of river water. Geochim. Cosmochim. Acta 56, 2099-2111. (doi:10.1016/0016-7037(92) 90332-D)
- 92. Angino EE, Billings GK, Andersen N. 1966 Observed variations in the strontium concentration of sea water. Chem. Geol. 1, 145-153. (doi:10.1016/0009-2541(66)90013-1)
- 93. Limburg K. 1995 Otolith strontium traces environmental history of subyearling American shad Alosa sapidissima. Mar. Ecol. Prog. Ser. 119, 25-35. (doi:10.3354/meps119025)

- 94. Howland KL, Tonn WM, Babaluk JA, Tallman RF. 2001 Identification of freshwater and anadromous inconnu in the Mackenzie river system by analysis of otolith strontium. Trans. Am. Fish. Soc. 130, 725-741. (doi:10.1577/1548-8659 (2001)130<0725:lofaai>2.0.Co;2)
- 95. Walther BD, Dempster T, Letnic M, McCulloch MT. 2011 Movements of diadromous fish in large unregulated tropical rivers inferred from geochemical tracers. PLoS ONE 6, e18351. (doi:10.1371/iournal.pone.0018351)
- 96 McCulloch M, Cappo M, Aumend J, Müller W. 2005 Tracing the life history of individual barramundi using laser ablation MC-ICP-MS Sr-isotopic and Sr/Ba ratios in otoliths. Mar. Freshw. Res. 56, 637-644. (doi:10.1071/MF04184)
- 97. Palace VP, Halden NM, Yang P, Evans RE, Sterling G. 2007 Determining residence patterns of rainbow trout using laser ablation inductively coupled plasma mass spectrometry (LA-ICP-MS) analysis of selenium in otoliths. Environ. Sci. Technol. 41, 3679-3683. (doi:10.1021/es0628093)
- 98. Lochet A, Limburg KE, Rudstam L, Montesdeoca M. 2010 Selenium incorporation in fish otoliths: effects of selenium and mercury from the water. Can. J. Fish. Aquat. Sci. 67, 1388–1397. (doi:10.1139/F10-070)
- 99. Yee H, Measures C, Edmond J. 1987 Selenium in the tributaries of the Orinoco in Venezuela, Nature 326, 686-689. (doi:10.1038/326686a0)
- 100. Sebastián MS, Armstrong B, Cordoba J, Stephens C. 2001 Exposures and cancer incidence near oil fields in the Amazon basin of Ecuador. Occup. Environ. Med. 58, 517-522. (doi:10.1136/oem.58.8.517)
- 101. Hurtig A-K, San Sebastián M. 2002 Geographical differences in cancer incidence in the Amazon basin of Ecuador in relation to residence near oil fields. Int. J. Epidemiol. 31, 1021–1027. (doi:10.1093/ije/31.5.1021)
- 102. Sebastián MS, Armstrong B, Stephens C. 2002 Outcomes of pregnancy among women living in

the proximity of oil fields in the Amazon basin of Ecuador. Int. J. Occup. Environ. Health 8, 312-319. (doi:10.1179/oeh.2002.8.4.312)

- 103. Hurtig A-K, San Sebastián M. 2004 Incidence of childhood leukemia and oil exploitation in the Amazon basin of Ecuador. Int. J. Occup. Env. Heal. 10, 245–250. (doi:10.1179/oeh.2004.10.3.245)
- 104. Sebastián MS, Hurtig AK. 2005 Oil development and health in the Amazon basin of Ecuador: the popular epidemiology process. Soc. Sci. Med. 60, 799-807. (doi:10.1016/j.socscimed.2004.06.016)
- 105. Lago Agrio Judgment. 2011 Nicolas Zambrano Lozada: Sucumbios Provincial Court of Justice.
- 106. Darley HC, Gray GR. 1988 Drilling fluid components. In Composition and properties of drilling and completion fluids. (eds HC Darley, GR Gray), pp. 535-616. Houston, TX: Gulf Professional Publishing.
- 107. Neff JM. 2008 Estimation of bioavailability of metals from drilling mud barite. Integr. Environ. Assess. Manage. 4, 184-193. (doi:10.1897/IEAM_2007-037.1)
- 108. Ulrich GA, Breit GN, Cozzarelli IM, Suflita JM. 2003 Sources of sulfate supporting anaerobic metabolism in a contaminated aquifer. Environ. Sci. Technol. 37, 1093-1099. (doi:10.1021/es011288a)
- 109. Karnachuk O, Kurochkina S, Tuovinen O. 2002 Growth of sulfate-reducing bacteria with solid-phase electron acceptors. Appl. Microbiol. Biotechnol. 58, 482-486 (doi:10.1007/s00253-001-0914-3)
- 110. Phillips EJ, Landa ER, Kraemer T, Zielinski R. 2001 Sulfate-reducing bacteria release barium and radium from naturally occurring radioactive material in oil-field barite. Geomicrobiol. J. 18, 167-182. (doi:10.1080/01490450151143453)
- 111. Winemiller K et al. 2016 Balancing hydropower and biodiversity in the Amazon, Congo, and Mekong. Science 351, 128-129. (doi:10.1126/science.aac7082)

16